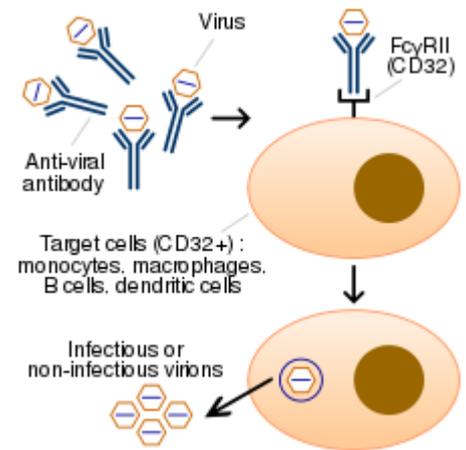


Antibody-dependent enhancement

Antibody-dependent enhancement (ADE), sometimes less precisely called **immune enhancement** or **disease enhancement**, is a phenomenon in which binding of a virus to suboptimal antibodies enhances its entry into host cells, followed by its replication.^{[1][2]} Antiviral antibodies promote viral infection of target immune cells by exploiting the phagocytic FcγR or complement pathway.^[3] After interaction with the virus the antibody binds Fc receptors (FcR) expressed on certain immune cells or some of the complement proteins. FcγR binds antibody via its fragment crystallizable region (Fc). Usually the process of phagocytosis is accompanied by the virus degradation, however, if the virus is not neutralized (either due to low affinity binding or targeting to a non-neutralizing epitope), antibody binding might result in a virus escape and therefore, enhanced infection. Thus, phagocytosis can cause viral replication, with the subsequent death of immune cells. The virus “deceives” the process of phagocytosis of immune cells and uses the host's antibodies as a Trojan horse. ADE may occur due to the non-neutralizing characteristic of the antibody, which bind viral epitopes other than those involved in a host cell attachment and entry. ADE may also happen due to the presence of sub-neutralizing concentrations of antibodies (binding to viral epitopes below the threshold for neutralization).^[4] In addition ADE can be induced when the strength of antibody-antigen interaction is below the certain threshold.^{[5][6]} This phenomenon might lead to both increased virus infectivity and virulence. The viruses that can cause ADE frequently share some common features such as antigenic diversity, abilities to replicate and establish persistence in immune cells.^[1] ADE can occur during the development of a primary or secondary viral infection, as well as after vaccination with a subsequent virus challenge.^{[1][7][8]} It has been observed mainly with positive-strand RNA viruses. Among them are Flaviviruses such as Dengue virus,^[9] Yellow fever virus, Zika virus,^{[10][11]} Coronaviruses, including alpha- and betacoronaviruses,^[12] Orthomyxoviruses such as influenza,^[13] Retroviruses such as HIV,^{[14][15][16]} and Orthopneumoviruses such as RSV.^{[17][18][19]}

The mechanism that involves phagocytosis of immune complexes via FcγRII / CD32 receptor is better understood compared to the complement receptor pathway.^{[20][21][22]} Cells that express this receptor are represented by monocytes, macrophages, some categories of dendritic cells and B-cells. ADE is mainly mediated by IgG antibodies,^[21] however, IgM along with complement,^[23] and IgA antibodies^{[15][16]} have also been shown to be trigger ADE.

ADE may cause enhanced respiratory disease and acute lung injury after respiratory virus infection (ERD) with symptoms of monocytic infiltration and an excess of eosinophils in respiratory tract.^[24] ADE along with type 2 T helper cell-dependent mechanisms may contribute to a development of the vaccine associated disease enhancement (VADE), which is not limited to respiratory disease.^[24] Some vaccine candidates that targeted coronaviruses, RSV virus and Dengue virus elicited VADE, and were terminated from further development or became approved for use only for patients who have had those viruses before.



In antibody-dependent enhancement, sub-optimal antibodies (the blue Y-shaped structures in the graphic) bind to both viruses and Fc gamma receptors (labeled FcγRII) expressed on immune cells promoting infection of these cells.

Contents

Coronavirus

Influenza

Dengue

HIV-1

Mechanism

Different virus serotypes

Conclusion

See also

References

Coronavirus

ADE was a concern during late clinical stages of vaccine development against COVID-19.^{[25][26]}

ADE was observed in animal studies during the development of coronavirus vaccines, but as of 14 December 2020 no incidents had been observed in human trials. "Overall, while ADE is a theoretical possibility with a COVID-19 vaccine, clinical trials in people so far have not shown that participants who received the vaccine have a higher rate of severe illness compared to participants who did not receive the vaccine."^{[27][28]}

Influenza

Prior receipt of 2008–09 TIV (Trivalent Inactivated Influenza Vaccine) was associated with an increased risk of medically attended pH1N1 illness during the spring-summer 2009 in Canada. The occurrence of bias (selection, information) or confounding cannot be ruled out. Further experimental and epidemiological assessment is warranted. Possible biological mechanisms and immunoepidemiologic implications are considered.^[29]

Natural infection and the attenuated vaccine induce antibodies that enhance the uptake of the homologous virus and H1N1 virus isolated several years later, demonstrating that a primary influenza A virus infection results in the induction of infection enhancing antibodies.^[30]

ADE was suspected in infections with influenza A virus subtype H7N9, but knowledge is limited.

Dengue

The most widely known ADE example occurs with dengue virus.^[31] Dengue is a single-stranded positive-polarity RNA virus of the family *Flaviviridae*. It causes disease of varying severity in humans, from dengue fever (DF), which is usually self-limited, to dengue hemorrhagic fever and dengue shock syndrome, either of which may be life-threatening.^[32] It is estimated that as many as 390 million individuals contract dengue annually.^[33]

ADE may follow when a person who has previously been infected with one serotype becomes infected months or years later with a different serotype, producing higher viremia than in first-time infections. Accordingly, while primary (first) infections cause mostly minor disease (dengue fever) in children, re-infection is more likely to be associated with dengue hemorrhagic fever and/or dengue shock syndrome in both children and adults.^[34]

Dengue encompasses four antigenically different serotypes (dengue virus 1–4).^[35] In 2013 a fifth serotype was reported.^[36] Infection induces the production of neutralizing homotypic immunoglobulin G (IgG) antibodies that provide lifelong immunity against the infecting serotype. Infection with dengue virus also produces some degree of cross-protective immunity against the other three serotypes.^[37] Neutralizing heterotypic (cross-reactive) IgG antibodies are responsible for this cross-protective immunity, which typically persists for a period of months to a few years. These heterotypic titers decrease over long time periods (4 to 20 years).^[38] While heterotypic titers decrease, homotypic IgG antibody titers increase over long time periods. This could be due to the preferential survival of long-lived memory B cells producing homotypic antibodies.^[38]

In addition to neutralizing heterotypic antibodies, infection can also induce heterotypic antibodies that neutralize the virus only partially or not at all.^[39] The production of such cross-reactive, but non-neutralizing antibodies could enable severe secondary infections. By binding to but not neutralizing the virus, these antibodies cause it to behave as a "trojan horse",^{[40][41][42]} where it is delivered into the wrong compartment of dendritic cells that have ingested the virus for destruction.^{[43][44]} Once inside the white blood cell, the virus replicates undetected, eventually generating high virus titers and severe disease.^[45]

A study conducted by Modhiran et al.^[46] attempted to explain how non-neutralizing antibodies down-regulate the immune response in the host cell through the Toll-like receptor signaling pathway. Toll-like receptors are known to recognize extra- and intracellular viral particles and to be a major basis of the cytokines production. In vitro experiments showed that the inflammatory cytokines and type 1 interferon production were reduced when the ADE-dengue virus complex bound to the Fc receptor of THP-1 cells. This can be explained by both a decrease of Toll-like receptor production and a modification of its signaling pathway. On one hand, an unknown protein induced by the stimulated Fc receptor reduces Toll-like receptor transcription and translation, which reduces the capacity of the cell to detect viral proteins. On the other hand, many proteins (TRIF, TRAF6, TRAM, TIRAP, IKKα, TAB1, TAB2, NF-κB complex) involved in the Toll-like receptor signaling pathway are down-regulated, which led to a decrease in cytokine production. Two of them, TRIF and TRAF6, are respectively down-regulated by 2 proteins SARM and TANK up-regulated by the stimulated Fc receptors.

One example occurred in Cuba, lasting from 1977 to 1979. The infecting serotype was dengue virus-1. This epidemic was followed by outbreaks in 1981 and 1997. In those outbreaks; dengue virus-2 was the infecting serotype. 205 cases of dengue hemorrhagic fever and dengue shock syndrome occurred during the 1997 outbreak, all in people older than 15 years. All but three of these cases were demonstrated to have been previously infected by dengue virus-1 during the first outbreak.^[47] Furthermore, people with secondary infections with dengue virus-2 in 1997 had a 3-4 fold increased probability of developing severe disease than those with secondary infections with dengue virus-2 in 1981.^[38] This scenario can be explained by the presence of sufficient neutralizing heterotypic IgG antibodies in 1981, whose titers had decreased by 1997 to the point where they no longer provided significant cross-protective immunity.

HIV-1

ADE of infection has also been reported in HIV. Like dengue virus, non-neutralizing level of antibodies have been found to enhance the viral infection through interactions of the complement system and receptors.^[48] The increase in infection has been reported to be over 350 fold which is comparable to ADE in other viruses like dengue virus.^[48] ADE in HIV can be complement-mediated or Fc receptor-mediated. Complements in the presence of HIV-1 positive sera have been found to enhance the infection of MT-2 T-cell line. The Fc-receptor mediated enhancement was reported when HIV infection was enhanced by sera from HIV-1 positive guinea pig enhanced the infection of peripheral blood mononuclear cells without the presence of any complements.^[49] Complement component receptors CR2, CR3 and CR4 have been found to mediate this Complement-mediated

enhancement of infection.^{[48][50]} The infection of HIV-1 leads to activation of complements. Fragments of these complements can assist viruses with infection by facilitating viral interactions with host cells that express complement receptors.^[51] The deposition of complement on the virus brings the gp120 protein close to CD4 molecules on the surface of the cells, thus leading to facilitated viral entry.^[51] Viruses pre-exposed to non-neutralizing complement system have also been found to enhance infections in interdigitating dendritic cells. Opsonized viruses have not only shown enhanced entry but also favorable signaling cascades for HIV replication in interdigitating dendritic cells.^[52]

HIV-1 has also shown enhancement of infection in HT-29 cells when the viruses were pre-opsonized with complements C3 and C9 in seminal fluid. This enhanced rate of infection was almost 2 times greater than infection of HT-29 cells with virus alone.^[53] Subramanian *et al.*, reported that almost 72% of serum samples out of 39 HIV positive individuals contained complements that were known to enhance the infection. They also suggested that the presence of neutralizing antibody or antibody-dependent cellular cytotoxicity-mediating antibodies in the serum contains infection-enhancing antibodies.^[54] The balance between the neutralizing antibodies and infection-enhancing antibodies changes as the disease progresses. During advanced stages of the disease the proportion of infection-enhancing antibodies are generally higher than neutralizing antibodies.^[55] Increase in viral protein synthesis and RNA production have been reported to occur during the complement-mediated enhancement of infection. Cells that are challenged with non-neutralizing levels of complements have been found have accelerated release of reverse transcriptase and the viral progeny.^[56] The interaction of anti-HIV antibodies with non-neutralizing complement exposed viruses also aid in binding of the virus and the erythrocytes which can lead to more efficient delivery of viruses to the immune-compromised organs.^[50]

ADE in HIV has raised questions about the risk of infections to volunteers who have taken sub-neutralizing levels of vaccine just like any other viruses that exhibit ADE. Gilbert *et al.*, in 2005 reported that there was no ADE of infection when they used rgp120 vaccine in phase 1 and 2 trials.^[57] It has been emphasized that much research needs to be done in the field of the immune response to HIV-1, information from these studies can be used to produce a more effective vaccine.

Mechanism

Interaction of a virus with antibodies must prevent the virus from attaching to the host cell entry receptors. However, instead of preventing infection of the host cell, this process can facilitate viral infection of immune cells, causing ADE.^{[1][3]} After binding the virus, the antibody interacts with Fc or complement receptors expressed on certain immune cells. These receptors promote virus-antibody internalization by the immune cells, which should be followed by the virus destruction. However, the virus might escape the antibody complex and start its replication cycle inside the immune cell avoiding the degradation.^{[3][23]} This happens if the virus is bound to low affinity antibody.

Different virus serotypes

There are several possibilities to explain the phenomenon of enhancing intracellular virus survival:

1) Antibodies against a virus of one serotype binds to a virus of a different serotype. The binding is meant to neutralize the virus from attaching to the host cell, but the virus-antibody complex also binds to the Fc-region antibody receptor (FcγR) on the immune cell. The cell internalizes the virus for programmed destruction but the virus avoids it and starts its replication cycle instead.^[58]

2) Antibodies against a virus of one serotype binds to a virus of a different serotype, activating the classical pathway of the complement system. The complement cascade system binds C1q complex attached to the virus surface protein via the antibodies, which in turn bind C1q receptor found on

cells, bringing the virus and the cell close enough for a specific virus receptor to bind the virus, beginning infection.^[23] This mechanism has been shown for Ebola virus *in vitro*^[59] and some flaviviruses *in vivo*.^[23]

Conclusion

When an antibody to a virus is unable to neutralize the virus it is forming sub-neutralizing virus-antibody complexes. Upon phagocytosis by macrophages or other immune cell such complex may release the virus due to poor binding with antibody. This happens during the step of acidification of phagosome^{[60][61]} before fusion with lysosome.^[62] The escaped virus begins its replication cycle within the cell, triggering ADE.^{[1][3][4]}

See also

- Original antigenic sin
- Other ways in which antibodies can (unusually) make an infection worse instead of better
 - Blocking antibody, which can be either good or bad, depending on circumstances
 - Hook effect, most relevant to in vitro tests but known to have some in vivo relevances

References

1. Tirado SM, Yoon KJ (2003). "Antibody-dependent enhancement of virus infection and disease". *Viral Immunology*. **16** (1): 69–86. doi:10.1089/088282403763635465 (https://doi.org/10.1089%2F088282403763635465). PMID 12725690 (https://pubmed.ncbi.nlm.nih.gov/12725690).
2. Wilder-Smith A, Hombach J, Ferguson N, Selgelid M, O'Brien K, Vannice K, et al. (January 2019). "Deliberations of the Strategic Advisory Group of Experts on Immunization on the use of CYD-TDV dengue vaccine" (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7119964). *The Lancet Infectious Diseases*. **19** (1): e31–e38. doi:10.1007/978-981-15-1045-8_2 (https://doi.org/10.1007%2F978-981-15-1045-8_2). ISBN 978-981-15-1044-1. PMC 7119964 (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7119964). PMID 30195995 (https://pubmed.ncbi.nlm.nih.gov/30195995).
3. Bournazos S, Gupta A, Ravetch JV (October 2020). "The role of IgG Fc receptors in antibody-dependent enhancement" (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7418887). *Nature Reviews Immunology*. **20** (10): 633–643. doi:10.1038/s41577-020-00410-0 (https://doi.org/10.1038%2Fs41577-020-00410-0). PMC 7418887 (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7418887). PMID 32782358 (https://pubmed.ncbi.nlm.nih.gov/32782358). S2CID 221108413 (https://api.semanticscholar.org/CorpusID:221108413).
4. Kulkarni, Ruta (2019-11-05). "Antibody-Dependent Enhancement of Viral Infections" (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7119964). *Dynamics of Immune Activation in Viral Diseases*: 9–41. doi:10.1007/978-981-15-1045-8_2 (https://doi.org/10.1007%2F978-981-15-1045-8_2). ISBN 978-981-15-1044-1. PMC 7119964 (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7119964).
5. Iwasaki A, Yang Y (June 2020). "The potential danger of suboptimal antibody responses in COVID-19" (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7187142). *Nature Reviews Immunology*. **20** (6): 339–341. doi:10.1038/s41577-020-0321-6 (https://doi.org/10.1038%2Fs41577-020-0321-6). PMC 7187142 (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7187142). PMID 32317716 (https://pubmed.ncbi.nlm.nih.gov/32317716).
6. Ricke D, Malone RW (2020). "Medical Countermeasures Analysis of 2019-nCoV and Vaccine Risks for Antibody-Dependent Enhancement (ADE)". *SSRN Working Paper Series*. doi:10.2139/ssrn.3546070 (https://doi.org/10.2139%2Fssrn.3546070). ISSN 1556-5068 (https://www.worldcat.org/issn/1556-5068). S2CID 216395996 (https://api.semanticscholar.org/CorpusID:216395996).

7. Tay MZ, Wiehe K, Pollara J (2019). "Antibody-Dependent Cellular Phagocytosis in Antiviral Immune Responses" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6404786>). *Frontiers in Immunology*. **10**: 332. doi:10.3389/fimmu.2019.00332 (<https://doi.org/10.3389%2Ffimmu.2019.00332>). PMC 6404786 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6404786>). PMID 30873178 (<https://pubmed.ncbi.nlm.nih.gov/30873178>).
8. Smatti, Maria K.; Al Thani, Asmaa A.; Yassine, Hadi M. (2018-12-05). "Viral-Induced Enhanced Disease Illness" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6290032>). *Frontiers in Microbiology*. **9**: 2991. doi:10.3389/fmicb.2018.02991 (<https://doi.org/10.3389%2Ffmicb.2018.02991>). ISSN 1664-302X (<https://www.worldcat.org/issn/1664-302X>). PMC 6290032 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6290032>). PMID 30568643 (<https://pubmed.ncbi.nlm.nih.gov/30568643>).
9. de Alwis R, Williams KL, Schmid MA, Lai CY, Patel B, Smith SA, et al. (October 2014). "Dengue viruses are enhanced by distinct populations of serotype cross-reactive antibodies in human immune sera" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4183589>). *PLOS Pathogens*. **10** (10): e1004386. doi:10.1371/journal.ppat.1004386 (<https://doi.org/10.1371%2Fjournal.ppat.1004386>). PMC 4183589 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4183589>). PMID 25275316 (<https://pubmed.ncbi.nlm.nih.gov/25275316>).
10. Khandia R, Munjal A, Dhama K, Karthik K, Tiwari R, Malik YS, et al. (2018). "Modulation of Dengue/Zika Virus Pathogenicity by Antibody-Dependent Enhancement and Strategies to Protect Against Enhancement in Zika Virus Infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5925603>). *Frontiers in Immunology*. **9**: 597. doi:10.3389/fimmu.2018.00597 (<https://doi.org/10.3389%2Ffimmu.2018.00597>). PMC 5925603 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5925603>). PMID 29740424 (<https://pubmed.ncbi.nlm.nih.gov/29740424>).
11. Plotkin S, Orenstein W (2012). "Yellow fever vaccine". *Vaccines* (6 ed.). Amsterdam: Elsevier. pp. 870–968. ISBN 9781455700905.
12. Yip MS, Leung NH, Cheung CY, Li PH, Lee HH, Daëron M, et al. (May 2014). "Antibody-dependent infection of human macrophages by severe acute respiratory syndrome coronavirus" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4018502>). *Virology Journal*. **11** (1): 82. doi:10.1186/1743-422X-11-82 (<https://doi.org/10.1186%2F1743-422X-11-82>). PMC 4018502 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4018502>). PMID 24885320 (<https://pubmed.ncbi.nlm.nih.gov/24885320>).
13. Winarski KL, Tang J, Klenow L, Lee J, Coyle EM, Manischewitz J, et al. (July 2019). "Antibody-dependent enhancement of influenza disease promoted by increase in hemagglutinin stem flexibility and virus fusion kinetics" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6660725>). *Proceedings of the National Academy of Sciences of the United States of America*. **116** (30): 15194–15199. doi:10.1073/pnas.1821317116 (<https://doi.org/10.1073%2Fpnas.1821317116>). PMC 6660725 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6660725>). PMID 31296560 (<https://pubmed.ncbi.nlm.nih.gov/31296560>).
14. Füst G (1997). "Enhancing antibodies in HIV infection". *Parasitology*. **115** Suppl (7): S127-40. doi:10.1017/s0031182097001819 (<https://doi.org/10.1017%2Fs0031182097001819>). PMID 9571698 (<https://pubmed.ncbi.nlm.nih.gov/9571698>).
15. Janoff, E. N.; Wahl, S. M.; Thomas, K.; Smith, P. D. (September 1995). "Modulation of human immunodeficiency virus type 1 infection of human monocytes by IgA" (<https://pubmed.ncbi.nlm.nih.gov/7658082/>). *The Journal of Infectious Diseases*. **172** (3): 855–858. doi:10.1093/infdis/172.3.855 (<https://doi.org/10.1093%2Finfdi%2F172.3.855>). ISSN 0022-1899 (<https://www.worldcat.org/issn/0022-1899>). PMID 7658082 (<https://pubmed.ncbi.nlm.nih.gov/7658082/>).
16. Kozlowski, P. A.; Black, K. P.; Shen, L.; Jackson, S. (1995-06-01). "High prevalence of serum IgA HIV-1 infection-enhancing antibodies in HIV-infected persons. Masking by IgG" (<https://pubmed.ncbi.nlm.nih.gov/7751656/>). *Journal of Immunology*. **154** (11): 6163–6173. ISSN 0022-1767 (<https://www.worldcat.org/issn/0022-1767>). PMID 7751656 (<https://pubmed.ncbi.nlm.nih.gov/7751656/>).

17. van Erp EA, van Kasteren PB, Guichelaar T, Ahout IM, de Haan CA, Luytjes W, et al. (November 2017). "In Vitro Enhancement of Respiratory Syncytial Virus Infection by Maternal Antibodies Does Not Explain Disease Severity in Infants" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5640862>). *Journal of Virology*. **91** (21). doi:10.1128/JVI.00851-17 (<https://doi.org/10.1128%2FJVI.00851-17>). PMC 5640862 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5640862>). PMID 28794038 (<https://pubmed.ncbi.nlm.nih.gov/28794038>).
18. Osiowy C, Horne D, Anderson R (November 1994). "Antibody-dependent enhancement of respiratory syncytial virus infection by sera from young infants" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC368388>). *Clinical and Diagnostic Laboratory Immunology*. **1** (6): 670–7. doi:10.1128/CDLI.1.6.670-677.1994 (<https://doi.org/10.1128%2FCDLI.1.6.670-677.1994>). PMC 368388 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC368388>). PMID 8556519 (<https://pubmed.ncbi.nlm.nih.gov/8556519>).
19. Gimenez HB, Chisholm S, Dornan J, Cash P (May 1996). "Neutralizing and enhancing activities of human respiratory syncytial virus-specific antibodies" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC170331>). *Clinical and Diagnostic Laboratory Immunology*. **3** (3): 280–6. doi:10.1128/CDLI.3.3.280-286.1996 (<https://doi.org/10.1128%2FCDLI.3.3.280-286.1996>). PMC 170331 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC170331>). PMID 8705669 (<https://pubmed.ncbi.nlm.nih.gov/8705669>).
20. Porterfield, James S.; Cardosa, M. Jane (1984), Notkins, Abner Louis; Oldstone, Michael B. A. (eds.), "Host Range and Tissue Tropisms: Antibody-Dependent Mechanisms", *Concepts in Viral Pathogenesis*, New York, NY: Springer, pp. 117–122, doi:10.1007/978-1-4612-5250-4_17 (https://doi.org/10.1007%2F978-1-4612-5250-4_17), ISBN 978-1-4612-5250-4
21. Bournazos S, Gupta A, Ravetch JV (October 2020). "The role of IgG Fc receptors in antibody-dependent enhancement" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7418887>). *Nature Reviews Immunology*. **20** (10): 633–643. doi:10.1038/s41577-020-00410-0 (<https://doi.org/10.1038%2Fs41577-020-00410-0>). PMC 7418887 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7418887>). PMID 32782358 (<https://pubmed.ncbi.nlm.nih.gov/32782358>).
22. Takada A, Kawaoka Y (2003). "Antibody-dependent enhancement of viral infection: molecular mechanisms and in vivo implications". *Reviews in Medical Virology*. **13** (6): 387–98. doi:10.1002/rmv.405 (<https://doi.org/10.1002%2Frmv.405>). PMID 14625886 (<https://pubmed.ncbi.nlm.nih.gov/14625886>). S2CID 9755341 (<https://api.semanticscholar.org/CorpusID:9755341>).
23. Cardosa, M. J.; Porterfield, J. S.; Gordon, S. (1983-07-01). "Complement receptor mediates enhanced flavivirus replication in macrophages" (<https://pubmed.ncbi.nlm.nih.gov/6864163/>). *The Journal of Experimental Medicine*. **158** (1): 258–263. doi:10.1084/jem.158.1.258 (<https://doi.org/10.1084%2Fjem.158.1.258>). ISSN 0022-1007 (<https://www.worldcat.org/issn/0022-1007>). PMC 2187083 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2187083>). PMID 6864163 (<https://pubmed.ncbi.nlm.nih.gov/6864163>).
24. Su, Shan; Du, Lanying; Jiang, Shibo (2020-10-16). "Learning from the past: development of safe and effective COVID-19 vaccines" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7566580>). *Nature Reviews Microbiology*: 1–9. doi:10.1038/s41579-020-00462-y (<https://doi.org/10.1038%2Fs41579-020-00462-y>). ISSN 1740-1534 (<https://www.worldcat.org/issn/1740-1534>). PMC 7566580 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7566580>). PMID 33067570 (<https://pubmed.ncbi.nlm.nih.gov/33067570>).
25. Lambert PH, Ambrosino DM, Andersen SR, Baric RS, Black SB, Chen RT, et al. (June 2020). "Consensus summary report for CEPI/BC March 12-13, 2020 meeting: Assessment of risk of disease enhancement with COVID-19 vaccines" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7247514>). *Vaccine*. **38** (31): 4783–4791. doi:10.1016/j.vaccine.2020.05.064 (<https://doi.org/10.1016%2Fj.vaccine.2020.05.064>). PMC 7247514 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7247514>). PMID 32507409 (<https://pubmed.ncbi.nlm.nih.gov/32507409>).
26. Haynes BF, Corey L, Fernandes P, Gilbert PB, Hotez PJ, Rao S, Santos MR, Schuitemaker H, Watson M, Arvin A (2020-10-19). "Prospects for a safe COVID-19 vaccine" (<https://doi.org/10.1126%2Fscitranslmed.abe0948>). *Science Translational Medicine*. **12** (568): eabe0948. doi:10.1126/scitranslmed.abe0948 (<https://doi.org/10.1126%2Fscitranslmed.abe0948>). PMID 33077678 (<https://pubmed.ncbi.nlm.nih.gov/33077678>). S2CID 224809822 (<https://api.semanticscholar.org/CorpusID:224809822>).

27. Teoh F, ed. (27 November 2020). "No evidence that COVID-19 vaccines cause more severe disease; antibody-dependent enhancement has not been observed in clinical trials" (<https://healthfeedback.org/claimreview/no-evidence-that-covid-19-vaccines-cause-more-severe-disease-antibody-dependent-enhancement-has-not-been-observed-in-clinical-trials/>) (Fact check). Health Feedback.
28. Gorski DH (14 December 2020). "It was inevitable that antivaxxers would claim that COVID-19 vaccines make females infertile" (<https://sciencebasedmedicine.org/it-was-inevitable-that-antivaxxers-would-claim-that-covid-19-vaccines-make-females-infertile/>). Science-Based Medicine. "the overall petition ... spends far more verbiage on PCR, Sanger sequencing, and the not-unreasonable but thus far not observed concern about antibody-dependent enhancement (ADE) due to a vaccine"
29. Skowronski DM, De Serres G, Crowcroft NS, Janjua NZ, Boulianne N, Hottes TS, et al. (April 2010). "Association between the 2008-09 seasonal influenza vaccine and pandemic H1N1 illness during Spring-Summer 2009: four observational studies from Canada" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2850386>). *PLOS Medicine*. **7** (4): e1000258. doi:10.1371/journal.pmed.1000258 (<https://doi.org/10.1371%2Fjournal.pmed.1000258>). PMC 2850386 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2850386>). PMID 20386731 (<https://pubmed.ncbi.nlm.nih.gov/20386731>).
30. Gotoff R, Tamura M, Janus J, Thompson J, Wright P, Ennis FA (January 1994). "Primary influenza A virus infection induces cross-reactive antibodies that enhance uptake of virus into Fc receptor-bearing cells". *The Journal of Infectious Diseases*. **169** (1): 200–3. doi:10.1093/infdis/169.1.200 (<https://doi.org/10.1093%2Finfdis%2F169.1.200>). PMID 8277183 (<https://pubmed.ncbi.nlm.nih.gov/8277183>).
31. "Model of antibody-dependent enhancement of dengue infection | Learn Science at Scitable" (<https://www.nature.com/scitable/content/model-of-antibody-dependent-enhancement-of-dengue-22403433/>). *www.nature.com*. Retrieved 2021-05-20.
32. Boonnak K, Slike BM, Burgess TH, Mason RM, Wu SJ, Sun P, et al. (April 2008). "Role of dendritic cells in antibody-dependent enhancement of dengue virus infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2292981>). *Journal of Virology*. **82** (8): 3939–51. doi:10.1128/JVI.02484-07 (<https://doi.org/10.1128%2FJVI.02484-07>). PMC 2292981 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2292981>). PMID 18272578 (<https://pubmed.ncbi.nlm.nih.gov/18272578>).
33. Ambuel Y, Young G, Brewoo JN, Paykel J, Weisgrau KL, Rakasz EG, et al. (15 September 2014). "A rapid immunization strategy with a live-attenuated tetravalent dengue vaccine elicits protective neutralizing antibody responses in non-human primates" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4046319>). *Frontiers in Immunology*. **5** (2014): 263. doi:10.3389/fimmu.2014.00263 (<https://doi.org/10.3389%2Ffimmu.2014.00263>). PMC 4046319 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4046319>). PMID 24926294 (<https://pubmed.ncbi.nlm.nih.gov/24926294>).
34. Guzman MG, Vazquez S (December 2010). "The complexity of antibody-dependent enhancement of dengue virus infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3185591>). *Viruses*. **2** (12): 2649–62. doi:10.3390/v2122649 (<https://doi.org/10.3390%2Fv2122649>). PMC 3185591 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3185591>). PMID 21994635 (<https://pubmed.ncbi.nlm.nih.gov/21994635>).
35. King CA, Anderson R, Marshall JS (August 2002). "Dengue virus selectively induces human mast cell chemokine production" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC155122>). *Journal of Virology*. **76** (16): 8408–19. doi:10.1128/JVI.76.16.8408-8419.2002 (<https://doi.org/10.1128%2FJVI.76.16.8408-8419.2002>). PMC 155122 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC155122>). PMID 12134044 (<https://pubmed.ncbi.nlm.nih.gov/12134044>).
36. Normile D (October 2013). "Tropical medicine. Surprising new dengue virus throws a spanner in disease control efforts". *Science*. **342** (6157): 415. Bibcode:2013Sci...342..415N (<https://ui.adsabs.harvard.edu/abs/2013Sci...342..415N>). doi:10.1126/science.342.6157.415 (<https://doi.org/10.1126%2Fscience.342.6157.415>). PMID 24159024 (<https://pubmed.ncbi.nlm.nih.gov/24159024>).

37. Alvarez G, Piñeros JG, Tobón A, Ríos A, Maestre A, Blair S, Carmona-Fonseca J (October 2006). "Efficacy of three chloroquine-primaquine regimens for treatment of *Plasmodium vivax* malaria in Colombia" (<https://doi.org/10.4269%2Fajtmh.2006.75.605>). *The American Journal of Tropical Medicine and Hygiene*. **75** (4): 605–9. doi:10.4269/ajtmh.2006.75.605 (<https://doi.org/10.4269%2Fajtmh.2006.75.605>). PMID 17038680 (<https://pubmed.ncbi.nlm.nih.gov/17038680>).
38. Guzman MG, Alvarez M, Rodriguez-Roche R, Bernardo L, Montes T, Vazquez S, et al. (February 2007). "Neutralizing antibodies after infection with dengue 1 virus" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2725871>). *Emerging Infectious Diseases*. **13** (2): 282–6. doi:10.3201/eid1302.060539 (<https://doi.org/10.3201%2Feid1302.060539>). PMC 2725871 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2725871>). PMID 17479892 (<https://pubmed.ncbi.nlm.nih.gov/17479892>).
39. Goncalvez AP, Engle RE, St Claire M, Purcell RH, Lai CJ (May 2007). "Monoclonal antibody-mediated enhancement of dengue virus infection in vitro and in vivo and strategies for prevention" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1868655>). *Proceedings of the National Academy of Sciences of the United States of America*. **104** (22): 9422–7. Bibcode:2007PNAS..104.9422G (<https://ui.adsabs.harvard.edu/abs/2007PNAS..104.9422G>). doi:10.1073/pnas.0703498104 (<https://doi.org/10.1073%2Fpnas.0703498104>). PMC 1868655 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1868655>). PMID 17517625 (<https://pubmed.ncbi.nlm.nih.gov/17517625>).
40. Peluso R, Haase A, Stowring L, Edwards M, Ventura P (November 1985). "A Trojan Horse mechanism for the spread of visna virus in monocytes". *Virology*. **147** (1): 231–6. doi:10.1016/0042-6822(85)90246-6 (<https://doi.org/10.1016%2F0042-6822%2885%2990246-6>). PMID 2998068 (<https://pubmed.ncbi.nlm.nih.gov/2998068>).
41. Chen YC, Wang SY (October 2002). "Activation of terminally differentiated human monocytes/macrophages by dengue virus: productive infection, hierarchical production of innate cytokines and chemokines, and the synergistic effect of lipopolysaccharide" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC136495>). *Journal of Virology*. **76** (19): 9877–87. doi:10.1128/JVI.76.19.9877-9887.2002 (<https://doi.org/10.1128%2FJVI.76.19.9877-9887.2002>). PMC 136495 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC136495>). PMID 12208965 (<https://pubmed.ncbi.nlm.nih.gov/12208965>).
42. Witayathawornwong P (January 2005). "Fatal dengue encephalitis" (https://web.archive.org/web/20110724015621/http://www.tm.mahidol.ac.th/seameo/2005_36_1/33-3352.pdf) (PDF). *The Southeast Asian Journal of Tropical Medicine and Public Health*. **36** (1): 200–2. PMID 15906668 (<https://pubmed.ncbi.nlm.nih.gov/15906668>). Archived from the original (http://www.tm.mahidol.ac.th/seameo/2005_36_1/33-3352.pdf) (PDF) on 24 July 2011.
43. Rodenhuis-Zybert IA, Wilschut J, Smit JM (August 2010). "Dengue virus life cycle: viral and host factors modulating infectivity". *Cellular and Molecular Life Sciences*. **67** (16): 2773–86. doi:10.1007/s00018-010-0357-z (<https://doi.org/10.1007%2Fs00018-010-0357-z>). PMID 20372965 (<https://pubmed.ncbi.nlm.nih.gov/20372965>). S2CID 4232236 (<https://api.semanticscholar.org/CorpusID:4232236>).
44. Guzman MG, Halstead SB, Artsob H, Buchy P, Farrar J, Gubler DJ, et al. (December 2010). "Dengue: a continuing global threat" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4333201>). *Nature Reviews. Microbiology*. **8** (12 Suppl): S7-16. doi:10.1038/nrmicro2460 (<https://doi.org/10.1038%2Fnmicro2460>). PMC 4333201 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4333201>). PMID 21079655 (<https://pubmed.ncbi.nlm.nih.gov/21079655>).
45. Dejnirattisai W, Jumnainsong A, Onsirisakul N, Fitton P, Vasanawathana S, Limpitikul W, et al. (May 2010). "Cross-reacting antibodies enhance dengue virus infection in humans" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3837288>). *Science*. **328** (5979): 745–8. Bibcode:2010Sci...328..745D (<https://ui.adsabs.harvard.edu/abs/2010Sci...328..745D>). doi:10.1126/science.1185181 (<https://doi.org/10.1126%2Fscience.1185181>). PMC 3837288 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3837288>). PMID 20448183 (<https://pubmed.ncbi.nlm.nih.gov/20448183>).

46. Modhiran N, Kalayanarooj S, Ubol S (December 2010). "Subversion of innate defenses by the interplay between DENV and pre-existing enhancing antibodies: TLRs signaling collapse" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3006139>). *PLOS Neglected Tropical Diseases*. *PLOS ONE*. **4** (12): e924. doi:10.1371/journal.pntd.0000924 (<https://doi.org/10.1371%2Fjournal.pntd.0000924>). PMC 3006139 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3006139>). PMID 21200427 (<https://pubmed.ncbi.nlm.nih.gov/21200427>).
47. Guzman MG (2000). "Dr. Guzman et al. Respond to Dr. Vaughn" (<https://doi.org/10.1093%2Faje%2F152.9.804>). *American Journal of Epidemiology*. **152** (9): 804. doi:10.1093/aje/152.9.804 (<https://doi.org/10.1093%2Faje%2F152.9.804>).
48. Willey S, Aasa-Chapman MM, O'Farrell S, Pellegrino P, Williams I, Weiss RA, Neil SJ (March 2011). "Extensive complement-dependent enhancement of HIV-1 by autologous non-neutralising antibodies at early stages of infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3065417>). *Retrovirology*. **8**: 16. doi:10.1186/1742-4690-8-16 (<https://doi.org/10.1186%2F1742-4690-8-16>). PMC 3065417 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3065417>). PMID 21401915 (<https://pubmed.ncbi.nlm.nih.gov/21401915>).
49. Levy JA (2007). *HIV and the pathogenesis of AIDS*. Wiley-Blackwell. p. 247. ISBN 978-1-55581-393-2.
50. Yu Q, Yu R, Qin X (September 2010). "The good and evil of complement activation in HIV-1 infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4002684>). *Cellular & Molecular Immunology*. **7** (5): 334–40. doi:10.1038/cmi.2010.8 (<https://doi.org/10.1038%2Fcmi.2010.8>). PMC 4002684 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4002684>). PMID 20228834 (<https://pubmed.ncbi.nlm.nih.gov/20228834>).
51. Gras GS, Dormont D (January 1991). "Antibody-dependent and antibody-independent complement-mediated enhancement of human immunodeficiency virus type 1 infection in a human, Epstein-Barr virus-transformed B-lymphocytic cell line" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC240554>). *Journal of Virology*. **65** (1): 541–5. doi:10.1128/JVI.65.1.541-545.1991 (<https://doi.org/10.1128%2FJVI.65.1.541-545.1991>). PMC 240554 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC240554>). PMID 1845908 (<https://pubmed.ncbi.nlm.nih.gov/1845908>).
52. Bouhlal H, Chomont N, Réquena M, Nasreddine N, Saidi H, Legoff J, et al. (January 2007). "Opsonization of HIV with complement enhances infection of dendritic cells and viral transfer to CD4 T cells in a CR3 and DC-SIGN-dependent manner" (<https://doi.org/10.4049%2Fjimmunol.178.2.1086>). *Journal of Immunology*. **178** (2): 1086–95. doi:10.4049/jimmunol.178.2.1086 (<https://doi.org/10.4049%2Fjimmunol.178.2.1086>). PMID 17202372 (<https://pubmed.ncbi.nlm.nih.gov/17202372>).
53. Bouhlal H, Chomont N, Haeffner-Cavaillon N, Kazatchkine MD, Belec L, Hocini H (September 2002). "Opsonization of HIV-1 by semen complement enhances infection of human epithelial cells" (<https://doi.org/10.4049%2Fjimmunol.169.6.3301>). *Journal of Immunology*. **169** (6): 3301–6. doi:10.4049/jimmunol.169.6.3301 (<https://doi.org/10.4049%2Fjimmunol.169.6.3301>). PMID 12218150 (<https://pubmed.ncbi.nlm.nih.gov/12218150>).
54. Subbramanian RA, Xu J, Toma E, Morisset R, Cohen EA, Menezes J, Ahmad A (June 2002). "Comparison of human immunodeficiency virus (HIV)-specific infection-enhancing and -inhibiting antibodies in AIDS patients" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC130693>). *Journal of Clinical Microbiology*. **40** (6): 2141–6. doi:10.1128/JCM.40.6.2141-2146.2002 (<https://doi.org/10.1128%2FJCM.40.6.2141-2146.2002>). PMC 130693 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC130693>). PMID 12037078 (<https://pubmed.ncbi.nlm.nih.gov/12037078>).
55. Beck Z, Prohászka Z, Füst G (June 2008). "Traitors of the immune system-enhancing antibodies in HIV infection: their possible implication in HIV vaccine development" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7115406>). *Vaccine*. **26** (24): 3078–85. doi:10.1016/j.vaccine.2007.12.028 (<https://doi.org/10.1016%2Fj.vaccine.2007.12.028>). PMC 7115406 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7115406>). PMID 18241961 (<https://pubmed.ncbi.nlm.nih.gov/18241961>).
56. Robinson WE, Montefiori DC, Mitchell WM (April 1990). "Complement-mediated antibody-dependent enhancement of HIV-1 infection requires CD4 and complement receptors". *Virology*. **175** (2): 600–4. doi:10.1016/0042-6822(90)90449-2 (<https://doi.org/10.1016%2F0042-6822%2890%2990449-2>). PMID 2327077 (<https://pubmed.ncbi.nlm.nih.gov/2327077>).

57. Gilbert PB, Peterson ML, Follmann D, Hudgens MG, Francis DP, Gurwith M, et al. (March 2005). "Correlation between immunologic responses to a recombinant glycoprotein 120 vaccine and incidence of HIV-1 infection in a phase 3 HIV-1 preventive vaccine trial" (<https://doi.org/10.1086%2F428405>). *The Journal of Infectious Diseases*. **191** (5): 666–77. doi:10.1086/428405 (<https://doi.org/10.1086%2F428405>). PMID 15688279 (<https://pubmed.ncbi.nlm.nih.gov/15688279>).
58. Takada A, Kawaoka Y (2003). "Antibody-dependent enhancement of viral infection: molecular mechanisms and in vivo implications". *Reviews in Medical Virology*. **13** (6): 387–98. doi:10.1002/rmv.405 (<https://doi.org/10.1002%2Frmv.405>). PMID 14625886 (<https://pubmed.ncbi.nlm.nih.gov/14625886>). S2CID 9755341 (<https://api.semanticscholar.org/CorpusID:9755341>).
59. Takada A, Feldmann H, Ksiazek TG, Kawaoka Y (July 2003). "Antibody-dependent enhancement of Ebola virus infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC164833>). *Journal of Virology*. **77** (13): 7539–44. doi:10.1128/JVI.77.13.7539-7544.2003 (<https://doi.org/10.1128%2FJVI.77.13.7539-7544.2003>). PMC 164833 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC164833>). PMID 12805454 (<https://pubmed.ncbi.nlm.nih.gov/12805454>).
60. Kinchen, Jason M.; Ravichandran, Kodi S. (October 2008). "Phagosome maturation: going through the acid test" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2908392>). *Nature Reviews. Molecular Cell Biology*. **9** (10): 781–795. doi:10.1038/nrm2515 (<https://doi.org/10.1038%2Fnrm2515>). ISSN 1471-0072 (<https://www.worldcat.org/issn/1471-0072>). PMC 2908392 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2908392>). PMID 18813294 (<https://pubmed.ncbi.nlm.nih.gov/18813294>).
61. Yates, Robin M.; Hermetter, Albin; Russell, David G. (2005-03-09). "The Kinetics of Phagosome Maturation as a Function of Phagosome/Lysosome Fusion and Acquisition of Hydrolytic Activity" (<https://dx.doi.org/10.1111/j.1600-0854.2005.00284.x>). *Traffic*. **6** (5): 413–420. doi:10.1111/j.1600-0854.2005.00284.x (<https://doi.org/10.1111%2Fj.1600-0854.2005.00284.x>). ISSN 1398-9219 (<https://www.worldcat.org/issn/1398-9219>). PMID 15813751 (<https://pubmed.ncbi.nlm.nih.gov/15813751>). S2CID 24021382 (<https://api.semanticscholar.org/CorpusID:24021382>).
62. Ong, Eugenia Z.; Zhang, Summer L.; Tan, Hwee Cheng; Gan, Esther S.; Chan, Kuan Rong; Ooi, Eng Eong (2017-01-13). "Dengue virus compartmentalization during antibody-enhanced infection" (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5234037>). *Scientific Reports*. **7** (1): 40923. Bibcode:2017NatSR...740923O (<https://ui.adsabs.harvard.edu/abs/2017NatSR...740923O>). doi:10.1038/srep40923 (<https://doi.org/10.1038%2Fsrep40923>). ISSN 2045-2322 (<https://www.worldcat.org/issn/2045-2322>). PMC 5234037 (<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5234037>). PMID 28084461 (<https://pubmed.ncbi.nlm.nih.gov/28084461>).

Retrieved from "https://en.wikipedia.org/w/index.php?title=Antibody-dependent_enhancement&oldid=1024203517"

This page was last edited on 20 May 2021, at 18:36 (UTC).

Text is available under the Creative Commons Attribution-ShareAlike License; additional terms may apply. By using this site, you agree to the Terms of Use and Privacy Policy. Wikipedia® is a registered trademark of the Wikimedia Foundation, Inc., a non-profit organization.